



GOVERNMENT OF GUAM
DEPARTMENT OF PUBLIC HEALTH AND SOCIAL SERVICES
DIPATTAMTON SALUT PUBLEKO YAN SETBISION SUSIAT

GUAM PUBLIC HEALTH LABORATORY SPECIMEN GUIDELINES
ACUTE NEUROLOGICAL ILLNESS OF UNDETERMINED ETIOLOGY

<p>Case Definition</p>	<p>To be considered a case and for laboratory testing, a patient must meet ALL 4 of the following criteria:</p> <ol style="list-style-type: none"> 1. Patient \leq21 years of age, 2. Acute onset of focal limb weakness, 3. On or after August 1, 2014 AND 4. An MRI showing spinal cord lesion largely restricted to gray matter
<p>Specimen Submission Guidelines</p>	<ul style="list-style-type: none"> • Call the DPHSS at (671) 689-3942 for pre-approval, before collecting and sending specimens for suspected Acute Neurological Illness. <u>NO SPECIMEN WILL BE ACCEPTED AT WITHOUT PRIOR CONSULTATION.</u> • Once vetted/approved by DPHSS, complete the specimen submission form with patient’s name (last name, first name), date of birth, unique Identification (ID) number/specimen ID number, date of collection, time of collection, initial of collector. • Samples from each patient should be submitted with a hard copy of Page 2 of the completed Patient Summary form (http://wwwdev.cdc.gov/ncird/investigation/viral/sep2014.html#forms), page 2 contains the patient demographic and ID information), “Acute Neurologic Illness with Limb Weakness in Children of Undetermined Etiology.” • Fill out <u>COMPLETELY</u> the CDC Specimen Submission form 50.34 (attached). • Call the GPHL at (671) 300-9082 or (671) 300-9085 to inform staff of the specimen delivery. • Specimens will be sent to a CDC reference laboratory for testing. • Physicians/providers will be notified of the result by the GPHL Technologist once the final result is received from CDC.
<p>Specimen Requirements</p>	<p>I. For Patients that are currently hospitalized: Collect and submit EACH of the following:</p> <ul style="list-style-type: none"> • CSF specimen • Upper respiratory tract specimen(ranked by 1st preference, to last preference) <ul style="list-style-type: none"> - Nasopharyngeal swab>>>nasal swab>>>nasal wash/aspirate >>>oropharyngeal swab • Serum sample <ul style="list-style-type: none"> - Acute and convalescent, if possible • TWO stool specimens, in accordance with the recommendations for poliovirus testing in all patients with a compatible clinical picture

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	<p style="text-align: center;">- Whole stool>> rectal swab</p> <p>II. For Patients that have already been discharged from the hospital:</p> <ul style="list-style-type: none"> • If less than 30 days since the hospital admission date, send any stored specimens from the list above. If no stored specimen is available, collect new specimen following the list above. • If 30 days or more since the hospital admission date, send any stored specimens from the list above.
<p>Specimen Collection Guidelines For Hospitalized Or Discharged Patients</p>	<ol style="list-style-type: none"> 1. CSF – collect as early in illness as possible, preferably at time of first evaluation or admission. Collect in sterile container, send as much sample as available. 2. Upper Respiratory Tract Specimen <ul style="list-style-type: none"> • Nasopharyngeal Swab – collect as early in illness as possible, preferably at time of first evaluation or admission. Store in viral transport medium. Minimum volume = 1 ml. • Nasal Swab - collect as early in illness as possible, preferably at time of first evaluation or admission. Store in viral transport medium. Minimum volume = 1 ml. • Nasopharyngeal wash or Aspirate - collect as early in illness as possible, preferably at time of first evaluation or admission. Collect in sterile container. Minimum volume = 1 ml • Oropharyngeal swab - collect as early in illness as possible, preferably at time of first evaluation or admission. Store in viral transport medium. Minimum volume = 1 ml. <p style="text-align: center;">Note: For NP or OP swabs, use only sterile Dacron or rayon swabs with plastic shafts or if available, flocked swabs. DO NOT use calcium alginate swabs or swabs with wooden sticks.</p> 3. Serum – Paired acute and convalescent specimens are optimal. Single serum specimens are also acceptable. Minimum volume ≥ 0.5 ml. 4. Stool – two samples total, collected at least 24 hours apart, both collected as early in illness as possible and ideally within 14 days of illness onset. <ul style="list-style-type: none"> • Whole stool – collect in sterile container, no special medium required. Minimum volume ≥ 1 gram. • Rectal swab – place and store swab in viral transport medium.

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	Note: For rectal swabs, use only sterile Dacron or rayon swabs with plastic shafts or if available, flocked swabs. DO NOT use calcium alginate swabs or swabs with wooden sticks.
Specimen Collection Guidelines for Deceased Patients	<p>In the event of a decedent, send both of these specimens:</p> <ol style="list-style-type: none"> 1. Fresh frozen tissue – place directly on dry ice or liquid nitrogen, store at -70°C and ship on dry ice. Representative sections from brain/spinal cord (including gray and white matter), heart, lung, liver, kidney and other organs as available. 2. Formalin-fixed or formalin-fixed paraffin-embedded tissue- Avoid prolonged fixation, tissues should have been fixed in formalin for 3 days, then transferred to 100% ethanol. Store and ship at room temperature in carriers to prevent breakage.
Specimen Transport Guidelines	<p>Transport frozen specimens with cold packs (2-8 C) to maintain frozen state or with dry ice to GPHL, except for the pathology specimens.</p> <p>Deliver specimens to the GPHL no later than 4 PM, business day Mondays-Fridays.</p> <p>Frozen specimens will be sent to CDC on dry ice.</p>
Rejection Criteria	<ul style="list-style-type: none"> • Specimens sent to the GPHL without prior consultation and notice. • Specimens collected in calcium alginate swabs or swabs with wooden sticks. • Specimen quantity is insufficient to perform the test; • Specimen not packed as described in Specimen Submission guidelines • Specimen is not received frozen. • Unlabeled specimens, incomplete label on specimen (Refer to Specimen Submission Guidelines). • Illegible/ incomplete Submission form. • Specimen label does not match the Submission form.
Result Notification	<p>Laboratory reports will be forwarded to the Submitting Facility, DPHSS, and the BCDC Administrator via FAX.</p> <p>Any other request for copies of laboratory reports, apart from that stipulated above will not be accepted.</p>
Contact Information	<p>Alan Mallari, Microbiologist III, GPHL (671) 300-9080 or (671) 300-9081 alan.mallari@dphss.guam.gov</p> <p>Lea Nisay, Microbiologist II, GPHL (Alternate) (671) 300-9083 FAX (671) 300-9098 lea.nisay@dphss.guam.gov</p>

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(Cont.) Contact Information	Anne Marie Santos, GPHL, Central Laboratory Administrator (671) 300-9082 or (671) 300-9085 Annemarie.santos@dphss.guam.gov
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Attachment: CDC Specimen Submission Form 50.34

References:

1. DPHSS Medical Advisory: Acute Neurologic Illness of Undetermined Etiology, Oct. 24, 2014
2. CDC Specimen Collection Instructions